

Characterization of a Novel Thrombin-like Enzyme, Globlase from the Venom of *Gloydius blomhoffii* (Japanese Mamushi)

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Authors' contributions

This work was carried out in collaboration among all authors. Authors YK and TN designed the study and managed the literature searches. Author YK also performed the sequence analysis and wrote the first draft of the manuscript. Authors YT and SO performed purification, electrophoresis and enzyme assays. All authors read and approved the final manuscript.

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ABSTRACT

Aims: To elucidate the coagulation mechanisms of a novel clotting factor isolated from *Gloydius blomhoffii* venom, its hydrolytic activity on various substrates were examined. Furthermore the primary structure was determined and compared with the other snake venom components.

Methodology: A thrombin-like enzyme was isolated from the crude venom of *G. blomhoffii* by DE52 Cellulose and CM52 Cellulose column chromatography. Enzyme activity was measured by using synthetic substrates (arginine esters, MCA-substrates and 3-(Acyloxy)-4-nitrobenzoic acid). Effect on fibrinogen was detected with bovine and human fibrinogen. Isoelectric point and molecular mass were measured by polyacrylamide gel electrophoresis and MALDI-TOF-MS. Amino acid sequence was decided with a protein sequencer by analyzing enzymatically cleaved peptides.

Results: A clotting factor was found to be homologous as indicated by a single band on SDS-PAGE, and the final preparation was named as globlase. Molecular mass of this enzyme was determined to be 13,876.36 Da and the isoelectric point was 8.8. Globlase showed arginine ester hydrolytic activity, and specificity for substrates of thrombin. Proteolytic activity and phospholipase

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A2 (PLA2) activity were not detected. Complete amino acid sequence analysis indicated that the primary structure of globlase is similar to PLA2. However the aspartic acid which exists in the active site of PLA2 was found to be substituted by glutamine.

Conclusion: It was shown in our current investigation that globlase is a novel thrombin-like enzyme isolated from *G. blomhoffii* venom. It was revealed that this enzyme had structure unlike the serine-protease such as the other thrombin-like enzymes.

Keywords: *Gloydus blomhoffii*; thrombin-like enzyme; primary structure; phospholipase A2.

1. INTRODUCTION

Serine-proteases which affect blood coagulation system are widely distributed in the venoms of Viperidae [1-3]. Various thrombin-like proteases have been purified from Viperidae snake venoms, which catalyze the cleavage of specific bond(s) in the fibrinogen molecule [4-9]. In our previous study, the detailed process of fibrin clot formation by a thrombin-like enzyme, bilineobin, from *Agkistrodon bilineatus* venom was reported [8,10].

Gloydus blomhoffii (Japanese Mamushi, which was formerly classified in genus *Agkistrodon*), widely distribute in Japan and it has been reported that 1,000 cases of bites occur annually [11]. Thrombin-like enzyme and arginine ester hydrolase are reported as the components of *G. blomhoffii* venom [12]. However, little change in prothrombin time (PT), activated partial thromboplastin time (APTT), or fibrinogen levels is observed even in severe cases [13]. Though the only thrombin-like enzyme purified from *G. blomhoffii* venom is halystase, this enzyme does not induce clot formation from fibrinogen. Therefore, we examined the existence of the other thrombin-like enzyme in this venom.

2. MATERIALS AND METHODS

2.1 Materials

Lyophilized crude venom of *G. blomhoffii* was purchased from Japan Snake Institute (Gunma, Japan). DE52- and CM52-Cellulose were the products of Whatman Laboratory (England). Tosyl-L-arginine methyl ester (TAME) and other arginine esters, methylcoumarylamide (MCA) substrates were obtained from the Peptide Institute, Inc. (Osaka, Japan). Fibrinogen (bovine, human and other animal species), hide powder azure, azocollagen, *p*-amidinophenyl methanesulfonyl fluoride (*p*-APMSF), *N*-bromosuccinimide (NBS), benzamide hydrochloride and glycoprotein detection kit were purchased from Sigma-Aldrich. Chymotrypsin, endoproteinase Lys-C and endoproteinase Arg-C

were from Roche Diagnostics. Other chemicals used were analytical grade from commercial sources.

2.2 Isolation and Characterization of Globlase

Crude venom (500 mg) was applied to a DE52 Cellulose column (1.5×45 cm) equilibrated with 5 mM sodium acetate buffer, pH 7.2. The adsorbed fraction was eluted with the same buffer containing 0.5 M NaCl. The fraction possessing fibrinogen clotting activity was collected and applied to a CM52 Cellulose column (1.5×45 cm) equilibrated with 5 mM sodium acetate buffer, pH 7.2. Elution was performed with a linear gradient from 5 mM to 0.5 M sodium acetate in a total volume of 600 ml. The homogeneity of the purified sample was confirmed by SDS-PAGE and reversed phase HPLC (Develosil ODS-HG-5 column, 4.6×250 mm). The molecular weight of final preparation was estimated by MALDI-TOF-MS, and the isoelectric point was determined with isoelectric-focusing polyacrylamide gel electrophoresis. The detection of sugar moieties of glycoproteins was performed with the modified Periodic Acid-Schiff (PAS) methods according to the instruction of suppliers.

2.3 Enzyme Activity and Other Assays

Arginine ester hydrolytic activity was assayed using tosyl-L-arginine methyl ester (TAME), benzoyl-L-arginine ethyl ester (BAEE) and benzoyl-L-arginine methyl ester (BAME) as substrates. The quantity of substrate hydrolyzed was determined by the hydroxamate method of Roberts [14]. One unit was defined as the quantity of protein which hydrolyzed 1 μmol of substrate per minute. Methylcoumarylamide (MCA) substrates which were designed for various serine-proteinases were also used to measure the enzymatic activity of globlase. Release of 7-amino-4-methylcoumarin was determined fluorometrically at 380 nm (λ_{ex}) and 460 nm (λ_{em}) according to the method of Morita et al. [15].

Indirect hemolysis of rabbit erythrocytes was employed to determine phospholipase A₂ (PLA₂) activity [16]. 3-(Acyloxy)-4-nitrobenzoic acid was synthesized to measure PLA₂ activity [17], and the enzyme assay was performed by the method of Holzer and Mackessy [18]. Proteolytic activity was measured by using casein, hide powder azure and azocollagen as the substrate [19-21].

In vitro clotting activity was assayed with fibrinogen of various animal species. Enzyme solution (0.1 ml) was added to 0.9 ml of fibrinogen solution (0.2% in saline). The reaction mixture was incubated at 37°C and the time interval necessary for the first appearance of fibrin clot was recorded. The analysis of digested fibrinogen fragments were performed as follows. The mixture of fibrinogen and globlase were incubated at 37°C, and the reaction was terminated with 20% trichloroacetic acid. The supernatant was collected by centrifugation (10,000 r.p.m. for 15 min) and separated by Develosil ODS-HG-5 column (4.6×250 mm). Identification of peptides was carried out by protein sequencer described in 2.4.

2.4 MALDI-TOF-MS and Amino Acid Sequence Analysis

Purified sample by reversed-phase HPLC was lyophilized, and then dissolved in 0.1% trifluoroacetic acid. The solution was mixed with sinapinic acid, and the molecular mass was measured by MALDI-TOF system (Autoflex speed, Bruker Daltonics).

Reduced and carboxymethylated sample [22] was enzymatically cleaved by chymotrypsin, endoproteinase Lys-C and endoproteinase Arg-C according to the reaction condition of suppliers manual. The amino acid sequence of digested fragments were separated by reversed-phase HPLC and analyzed by Applied Biosystems 491 protein sequencer and Model 120A PTH analyzer.

3. RESULTS AND DISCUSSION

3.1 Isolation and Characterization of Globlase

3.1.1 Isolation of globlase

Crude venom was applied to a DE52 Cellulose column (Fig. 1A). Clotting and TAME hydrolytic activities were found in fractions 1 and 2. Fraction 1 was further purified by a CM-Cellulose column (Fig. 1B), and the enzyme activity was

found between the fraction number 160 and 170. This fraction was homogeneous with various tests such as SDS-PAGE (Fig. 1B-insert), isoelectric focusing and reversed-phase HPLC (Fig. 1C). This protein was named as "Globlase" (a thrombin-like enzyme from *G. blomhoffii*). By this procedure, 10.8 mg of purified preparation was obtained from 500 mg of crude venom.

3.1.2 Biochemical properties

The molecular mass of globlase measured by MALDI-TOF-MS was 13,876.36 Da with the isoelectric point of 8.8. The modified Periodic Acid-Schiff (PAS) method indicated that this protein did not contain any carbohydrates.

Globlase possessed coagulation activity on bovine and human fibrinogen. Enzyme specificity was measured by synthetic peptides TAME, BAME and BAEE for serine proteinase (Table 1). Among these substrates, TAME was hydrolyzed most actively and the specific activity was 27.04 units/mg of protein. *t*-Butoxycarbonyl-Asp(OBz)-Pro-Arg-MCA which was designed for α -thrombin was the most suitable substrate among various MCA-substrates which participate in blood coagulation. The hydrolyzing activity of globlase against MCA-substrates of factor Xa, activated protein C and plasmin were relatively low. Proteolytic activities on casein, hide powder azure and azocollagen were relatively low or not detected. Neither hydrolytic activity on a synthetic PLA₂ substrate nor indirect hemolytic activity was detected.

TAME hydrolytic activity and clotting activity of globlase (30 μ g) were stable to heat treatment. The residual activities after the heat treatment at 100°C for 10 min were 38.8% and 34.7%, respectively. Both activities were inhibited by *p*-APMSF and NBS. However, no inhibition was observed with benzamidine or EDTA.

According to the previous study, it is reported that halystase, a serine proteinase, exists in *G. blomhoffii* venom [9]. This enzyme consists of 238 amino acid residues and N-linked carbohydrates with the molecular mass of 38 kDa. Halystase is structurally similar to thrombin-like snake venom enzymes and cleaves fibrinogen. However, it does not induce fibrin clot formation. Since coagulation factor except halystase is not isolated from *G. blomhoffii* venom, globlase is considered a novel thrombin-like enzyme.

3.1.3 Effect of globlase on fibrinogen

Since the final preparation possessed clotting activity, the effect of globlase (32.5 μ g) on fibrinogen of various animal species was further tested. The clot formation of bovine fibrinogen was the fastest (30 sec), followed by sheep and human fibrinogen (45 and 70 sec, respectively). Fibrinogen from dog, horse, guinea pig was also coagulated, but their clotting time was relatively slow. Time-dependent degradation of bovine fibrinogen was observed on SDS-PAGE (Fig. 2A). A α chain was immediately hydrolyzed in a few minutes. It seems B β and γ chains of fibrinogen were resistant to hydrolysis with globlase.

Two major peaks were detected in the elution profile of the supernatant of digested bovine fibrinogen (Fig. 2B). The peptide eluted at 19 min was identified as fibrinopeptide A. However, the

peptide obtained in elution time 20-21min was not identified. These results indicate that globlase specifically hydrolyze A α chain of bovine fibrinogen and release fibrinopeptide A.

Various thrombin-like enzymes have been classified according to their ability to release fibrinopeptides [2]. Similar to thrombin, the enzymes from *Bitis gabonica* and *Bothrops insularis* catalyze fibrinopeptides A and B [23,24]. But most of venom thrombin-like enzymes release either fibrinopeptide A or B. Similar to globlase, reported fibrinopeptide A-releasing enzymes hydrolyze TAME and/or synthetic thrombin substrate. However, they belong to serine-proteinase family and the molecular weights are larger than globlase. Furthermore, most of these enzymes contain carbohydrate chains [2]. Therefore, globlase is supposed to be a novel thrombin-like enzyme.

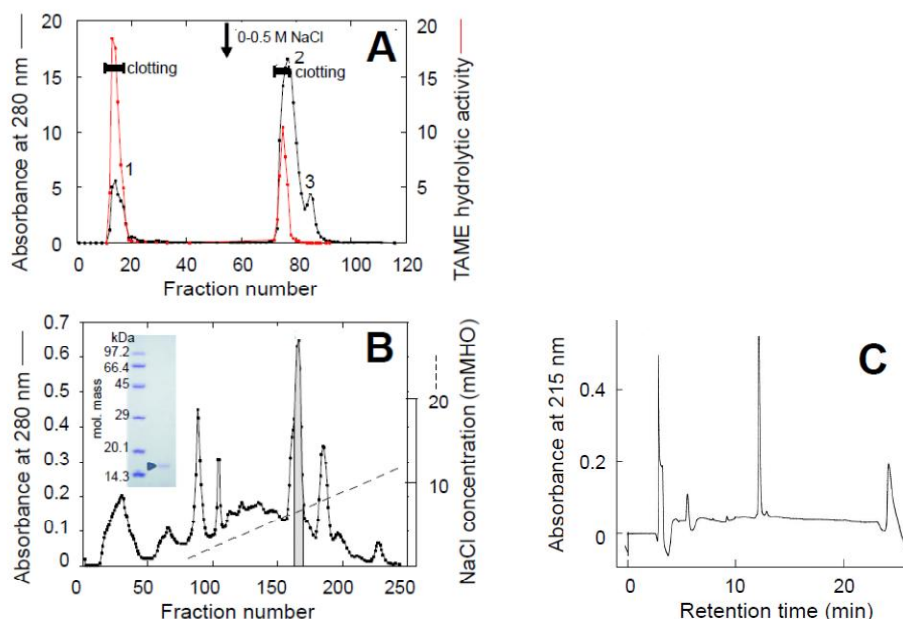


Fig. 1. Isolation of globlase

(A) The first step: Crude venom of *G. blomhoffii* (500 mg) was applied on a DE52 Cellulose column (1.5×45 cm) equilibrated with 5 mM sodium acetate. The adsorbed fraction was eluted with equilibration buffer containing 0.5 M NaCl

(B) The second step: Fraction 1 from the first step was applied to a CM52 Cellulose column (1.5×45 cm) equilibrated with 5 mM acetate buffer (pH 7.2). Elution was performed with a linear gradient from 5 mM to 0.5 M sodium acetate buffer (pH 7.2) in a total volume of 600 mL. Fractions of 3.0 mL were collected at a flow rate of 10.8 mL/hr. The fraction possessing TAME hydrolytic activity was indicated by shadowed area. (insert) SDS-PAGE of isolated enzyme

(C) Reversed-phase HPLC profile of the final preparation of globlase: The purified fraction from CM52 Cellulose column was applied to Develosil ODS-HG-5 column (4.5 × 250 mm) equilibrated with 0.1 % TFA in H₂O. Elution was performed with a linear gradient from 0 to 100 % in 0.1 % TFA in acetonitrile over 20 min

Table 1. Enzyme activities of globlase

Substrate	Activity
Arginine esters	(units/mg)
TAME (Tos [*] -L-Arg methyl ester)	27.04
BAME (Bz ^{**} -L-Arg methyl ester)	3.43
BAEE (Bz-L-Arg ethyl ester)	2.10
MCA [†] -substrates (designed for)	AMC ^{††} released (μmol/min/mg)
Boc ^{***} -Asp(OBz)-Pro-Arg-MCA (α-thrombin)	8.21
Boc-Val-Pro-Arg-MCA (α-thrombin)	2.71
Boc-Ile-Glu-Gly-Arg-MCA (factor Xa)	0.90
Boc-Leu-Ser-Thr-Arg-MCA (activated protein C)	1.20
Boc-Val-Leu-Lys-MCA (plasmin)	0.75
Proteinase	(units/mg)
Casein	ND
Hide powder azure	
Azocollagen	0.41
Phospholipase A ₂	(units/mg)
3-(Acyloxy)-4-nitrobenzoic acid	ND

*Tos: tosyl. **Bz: benzoyl. ***Boc: t-butoxybalbonyl. †MCA: Methylcoumarylamide.

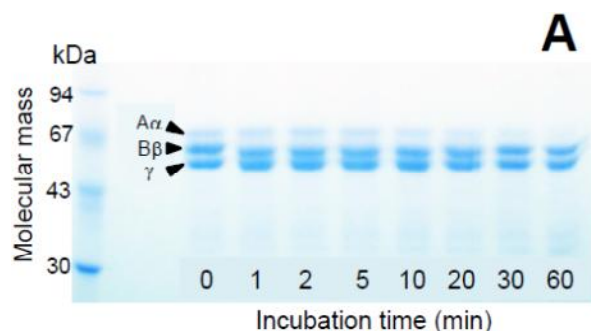
††AMC: aminomethylcoumarin. ND: not detected

3.1.4 Amino acid sequence of globlase

The sequence from the amino-terminal to His(47) was identified by the direct analysis of native globlase (Fig. 3). The sequence from Gln(48) to Cys(122) were determined by using the results of analysis of digested fragments. From these results, globlase was found to be a single chain protein consisting of 122 amino acid residues. The theoretical molecular weight calculated from the total sequence (Fig. 3) was 13876.97, and it consistent with the result obtained by MALDI-TOF-MS.

Since the primary structure of globlase seems to homologous with PLA₂, the sequence was compared with PLA₂s from *G. blomhoffii* venom [25,26] (Fig. 4). The enzymes from the venom of *G. ussuriensis* and *P. flavoviridis*, which inhabit the near geographical range, were also aligned [27,28]. The sequence of basic and acidic PLA₂s isolated from *G. blomhoffii* venom were apparently different from that of globlase, and the

identity of these enzymes was 91.0% and 60.7%, respectively. It is interesting that enzyme of *G. ussuriensis*, which inhabit the eastern part of Eurasian continent, is most homologous to globlase. The homology of both enzymes was 97.5%, and only the difference in three amino acid residues was found. Zhang et al. reported that Gln49-PLA₂ from *G. ussuriensis* venom possess thrombin-like activity [29]. Because the His residue of active site [30] replaced to Gln, this enzyme lacks PLA₂ activity. Furthermore, this enzyme hydrolyzed arginine ester and human fibrinogen, and its clotting activity was equivalent to 1100 NIH thrombin U/mg. Among snake venom PLA₂ analogues reported, enzyme possessing similar characteristics with thrombin-like serine proteinase is only globlase and Gln49-PLA₂. It is interesting that snakes inhabiting the geographically close area have the similar enzyme. Detailed examination of PLA₂ analogues without PLA₂ activity is necessary to find out the enzymes possessing similar characteristics to these two enzymes.



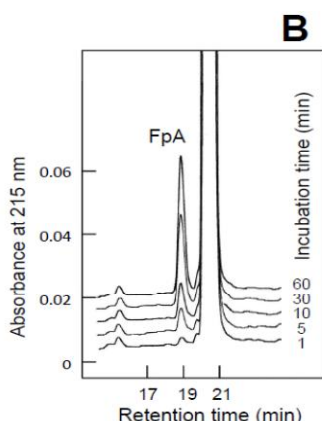


Fig. 2. Effect of globlase on bovine fibrinogen

(A) Time-dependent digestion of bovine fibrinogen. Fibrinogen (0.2 % in saline) was incubated with globlase at 37°C for various time intervals and analyzed by SDS-PAGE. (B) Elution profile of the digests of fibrinogen by reversed-phase HPLC. Bovine fibrinogen (5 mg) and globlase (1.63 μg) were incubated at 37°C for indicated time intervals. The reaction was terminated by addition of 20 % TCA, and centrifuged (10,000 r.p.m.) for 15 min. The supernatant was separated by Develosil ODS-HG-5 column (4.6×250 mm) with a linear gradient from 15 to 35 % acetonitrile in 40 min



Fig. 3. Determination of amino acid sequence of globlase

Arrows indicate residues determined by sequence analysis. The following abbreviations were used for the peptides digested by: K; endoproteinase Lys-C, C; chymotrypsin, and R; endoproteinase Arg-C

	1	10	20	30	
Blomhobin (<i>G. blomhoffii</i>)	S L L Q F R K M I K K M T G K E P V V A Y A F Y G	C Y C G S	G R G R K	◆	◆
basic PLA ₂ (<i>G. blomhoffii</i>)	H L L Q F R K M I K K M T G K E P V I S Y A F Y G	C Y C G S	G R G R K		
acidic PLA ₂ (<i>G. blomhoffii</i>)	S L M Q F E T L I M K I A G R S G I W Y Y G S Y G	C Y C G A	G G Q G R		
Gln49-PLA ₂ (<i>G. ussuriensis</i>)	S L L Q F R K M I K K M T G K E P V V S Y A F Y G	C Y C G S	G R G R K		
PLA-B (<i>P. flavoviridis</i>)	H L L Q F R K M I K K M T G K E P I V S Y A F Y G	C Y C G K	G R G R K		
	40	50	60	70	80
	P K D A T D R C C F V H Q C C Y E K V T G C D P K W D D Y T Y S W K D G D I V C G G D D P				
	P K D A T D R C C F V H D C C Y E K V T G C K P K W D D Y T Y S W K N G D I V C G G D D P				
	P Q D A S D R C C F V H D C C Y G K V T G C D P K L D V Y T Y T E E N G A I V C G G D D P				
	P K D A T D R C C F V H Q C C Y E K V T G C D P K W D D Y T Y S W K D G D I V C G G D D P				
	P K D A T D R C C F V H D C C Y E K V T G C D P K W D Y Y T Y S S E N G D I V C E G D N P				
	90	100	110	120	
	C K K E V C E C D R A A A I C F R D N L K T Y K K I Y M A Y P D I F C S S D P E K C				this study
	C K K E I C E C D R A A A I C F R D N L K T Y K K R Y M A Y P D I L C S S K S E K C				91.0%
	C K K Q I C E C D K D A A I C F R D N I D T Y D N K Y W F F P A K N C Q S E S E P C				60.7%
	C K K E V C E C D R A A A I C F R D N L K T Y K K I Y M A Y P D I F C S S K S E K C				97.5%
	C T K E V C E C D K A A A I C F R D N L K T Y K K R Y M T F P D I F C T D P T E K C				83.6%

Fig. 4. Comparison of amino acid sequence of globlase and phospholipase A₂S

The protein sequence was aligned based on the position of Cys residues. The important amino acids in the catalytic center of phospholipase A₂ are boxed. The calcium-binding site is indicated with (◆)

4. CONCLUSION

It was shown in our current investigation that globlase is a novel thrombin-like enzyme isolated from *G. blomhoffii* venom. It was revealed that this enzyme had unique structure unlike the other serine-proteases. This result is interesting in studying the process of the evolution of snake enzymes.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

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