



Beneficial Effects of Eucalyptol in the Pathophysiological Changes of the Respiratory System: A Proposal for Alternative Pharmacological Therapy for Individuals with COPD

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Authors' contributions

This work was carried out in collaboration between all authors. Authors FLG, GRS and IFMGN wrote the first draft of the manuscript and managed the literature searches. Author DSS reviewed and edited the manuscript. Author FSAC supervised the study. All authors read and approved the final manuscript.

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Review Article

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ABSTRACT

It is estimated that there will be an increase in the incidence of chronic obstructive pulmonary disease (COPD) in the coming decades. Thus, the pharmacological attributes of products of plant origin should be considered as an important economic and scientific strategy in the investigation of therapeutic alternatives, since their experimental validations are indispensable to substantiate the reliability of these products in the treatment of chronic diseases. Like biologically active compounds,

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Eucalyptol, also known as 1,8- cineole, is the major constituent of the leaf oil of eucalyptus species, such as *Eucalyptus globulus* and *Eucalyptus tereticornis*. It is a terpenoid oxide, free of steroid-like side effects. This study is based on a review of the specialised literature with purpose to discuss the biological effects of Eucalyptol in the respiratory system and its interaction with some of the most promising targets in the treatment of COPD, such as: receptors and membrane channels, oxidative stress, transcription and expression of cytokines, cell adhesion molecules and neutrophil chemotaxis, proteases and remodeling.

Keywords: *Anti-inflammatory; biological activity; COPD; eucalyptol; herbal medicine; respiratory system; 1,8- cineole.*

1. INTRODUCTION

Individuals with chronic physiologic dysfunctions such as cancer, diabetes, cardiovascular disease, asthma and chronic obstructive pulmonary disease (COPD) are often affected by a number of factors including irregular physical activity, poor eating habits, smoking, and environmental pollutants [1].

Although it is preventable and treatable, COPD is still the fourth leading cause of death in the world, and it is estimated that there will be an increase in its incidence in the coming decades due to population ageing and continuous exposure to its risk factors [2]. In parallel, the study of the pharmacological attributes of plant origin products used for medicinal purposes should be recognised as an important economic and scientific strategy in the investigation of therapeutic alternatives, since their experimental validations are indispensable to base the reliability of these products. With this motivation, components derived from plant species have been widely used in a wide variety of diseases, including chronic diseases [3].

Like biologically active compounds, Eucalyptol, also known as 1,8- cineole, is a major constituent of the leaf oil of eucalyptus species, such as *Eucalyptus globulus* Labill and *Eucalyptus tereticornis* SM. It is classified as a terpenoid oxide, compound responsible for fragrance and pleasant taste, endowed with an immense variety of structures and biological activities, free of steroid-like side effects. Thus, systemic therapy with Eucalyptol seems to be favourable in relation to its lipophilicity related to the terpene group, and its excretion predominant by exhalation [4-6].

Such characteristics of this compound attribute approval of Eucalyptol by the US Food and Drug Administration (USFDA) for consumption as a food additive and license as a medicinal product

(Soledum™ capsules, Cassella-med, Cologne, Germany) in Germany [7]. In view of the above, this review aims to describe the cell signalling pathways and biological activities of Eucalyptol in the respiratory system, to provide scientific support on its efficacy as an alternative therapy for the treatment of COPD.

This study is based on the reviews of the specialised literatures, in which references were collected from books and scientific articles selected from electronic databases such as Scielo, Medline, Pubmed and ScienceDirect. The inclusion criteria for the studies found were the therapeutic approaches in COPD, the biological activity of Eucalyptol on the respiratory system, as well as the cellular signalling pathways of this constituent. We excluded studies that reported aspects with an emphasis in another discussion that the focus was not related to the respiratory system or pharmacological properties of Eucalyptol.

2. GLOBAL INITIATIVE PROJECT ON COPD

It is estimated that there will be an increase in the incidence of COPD in the coming decades. Since 2001, the global strategy for the diagnosis, management, and prevention of COPD has been a valuable resource for professional health promoters. Thus, the Global Initiative for Chronic Obstructive Pulmonary Disease (GOLD) project strives to improve prevention and care in COPD worldwide. Its specific topics address diagnosis, management of exacerbations in Asthma and COPD, and means of treating the disease when in its stable stage [2].

Despite current and future needs, the scientific committee of the GOLD project, in its catalogue of scientific papers suggested for reading and deepening, explains the need for discussion, elucidation and new research on the pathophysiological factors involved in COPD, such as: immune system, membrane specific

receptors, gene transcription factors, cytokines, chemokines, proteases, antiproteases [8-14] and aggravating and/or causative agents of the disease, which cause dysfunction in the airways and pulmonary parenchyma, such as exposure to cigarette smoke and environmental pollutants [15-18].

A greater understanding of the inflammatory mechanisms involved in COPD, achieved in the last decades, has resulted in the identification of several processes and goals for the development of new anti-inflammatory treatments [19]. The following topics bring a sequential approach to the action of pathophysiological mediators parallel to the regulation of these exerted by Eucalyptol.

3. PHARMACOLOGICAL STUDIES

3.1 Receptors and Membrane Channels

Hypersecretion of mucus, one of the causes of airflow limitation in COPD, is due to the increase in the number of goblet cells and submucosal glands, both due to chronic irritation in the airways by noxious agents. In this situation, many mediators stimulate mucus hypersecretion exerting their effects through the activation of the epidermal growth factor receptor (EGFR), whose ligands, such as transforming growth factor alpha (TGF- α), are produced by neutrophils and macrophages [20-23].

Analyses were performed on bronchial biopsy specimens obtained from asthmatic individuals and patients with COPD. Results showed a positive correlation between EGFR and mucin MUC5AC expression [24,25], as EGFR acts as a transcription factor that plays a regulatory role in the expression of many genes important for inflammation [26,27]. Zhou and collaborators [28] performed a study to elucidate the anti-inflammatory mechanisms in monocytes obtained from asthmatic subjects incubated with Eucalyptol thirty minutes before being stimulated with lipopolysaccharides (LPS). In this work, it was observed that Eucalyptol in a concentration-dependent manner (1, 10, and 100 mg/L, 30 min) was able to inhibit EGFR synthesis, providing an evidence of the role of 1,8- cineole in the control of inflammation and limitation to airway flow.

In a study conducted by Nascimento and collaborators [29], 1,8- cineole reduced the tracheobronchial resistance *in vivo* after bronchospasm was induced by the challenge to

carbachol. A similar effect was seen when compared to the response obtained with fenoterol, a drug used in asthmatic crises and exacerbation of COPD. In addition, it also directly relaxed *in vitro* the airway smooth muscle previously contracted with the induction of carbachol, a high concentration of potassium and histamine. Inhibition of phasic contractions suggests that Eucalyptol has an antagonistic action on the transmembrane influx of calcium or its intracellular action as a second messenger.

Bastos and collaborators [30], in a model of airway hyper reactivity with subsequent treatment with a single dose of Eucalyptol (1 mg/mL) administered by inhalation, significantly developed lower tracheal ring contractions when compared to the untreated group. Specifically, we observed Eucalyptol's preferential action on voltage-operated calcium channels (VOCCs).

In accordance with such myorelaxant properties, Soares and collaborators [31] have shown that 1,8- cineol could induce a negative inotropic effect on rat heart tissues, while it blocked the influx of Ca^{2+} through the VOCCs located in the sarcolemma of cardiac myocytes. Therefore, the relaxation induced by Eucalyptol in the muscle tissue of the trachea and bronchus in the murine model may be related to its negative interference in the influx of calcium through the cell membrane.

Although it is still a question of transmembrane proteins, it is important to note that a signalling pathway associated with Toll-like 4 standard recognition receptors (TLR4), such as the activation of p38 mitogen-activated protein kinase (MAPK p38), play a critical role in inflammation allergic reaction [32]. Continuous inhalation of irritants, such as cigarette smoke, fossil fuel gases and environmental particles, activate TLR4 [33,34]. This mechanism leads to the propagation of an innate immune response, with activation of airway epithelial cells and secretion of mucus [35].

Zhao and collaborators [36], investigated the expression of these receptors in mice with LPS-induced lung inflammation after treatment with Eucalyptol. In this study, a single oral dose of Eucalyptol (100 mg/kg) was found to decrease TLR4 expression when compared to the non-constituent group and the positive control group treated with prednisone, a substance used in anti-inflammatory drugs. Later, the effects of Eucalyptol in a model of solution-induced asthma

composed of dust mites at home were investigated by Lee and collaborators [37], where TLR4 suppression and mitogen-activated protein kinase p38 (MAPK p38) in mice was treated with Eucalyptol (10 mg/mL), via nebulization, before each exposure to the aggressive agent.

3.2 Oxidative Stress

The redox imbalance is also an important mechanism of conduction in the pathophysiology of chronic diseases and a crucial target for therapies in COPD [38], because reactive oxygen species (ROS) activate nuclear factor kappa B (NF- κ B) and MAPK p38, thus leading to a further intensification of inflammatory genes and inhibition of the activity of endogenous antiproteases. This suggests that antioxidants may be very useful in the treatment of COPD by reducing the inflammatory process, as well as repairing and reversing resistance to corticosteroids [19].

The production of reactive oxygen species (ROS) caused by smoking is linked to the protease/antiprotease imbalance that contributes to the development of COPD [39,40]. Kennedy-Feitosa and collaborators [41], analysed the efficacy of Eucalyptol against acute lung inflammation caused by cigarette smoke (CF), in which mice were exposed to CF and treated with Eucalyptol (10 mg/mL) via inhalation 15 minutes a day, for 5 days. In this protocol, it was observed that the group treated with Eucalyptol, when compared to the group exposed to smoke and untreated, was able to reduce ROS levels, confirmed by the reduction of the enzymatic activities of catalase (CAT) and superoxide dismutase (SOD). In parallel, the compound reduced oxidative damage through lipid peroxidation, evidenced by reduced levels of malondialdehyde (MDA).

3.3 Transcription and Expression of Cytokines

Both MAPK p38 and oxidative stress induce the activation of NF- κ B by promoting the transcription of pro-inflammatory cytokines [42-45], resulting in its translocation to the nucleus, adhesion to DNA and effectuation of genetic transcription. The pathway of activation of this factor is associated with the transcription of genes involved in the inflammatory process, such as cytokines, chemokines and adhesion molecules [46].

In a model of acute lung injury (IPA) induced by LPS, BALB / C, mice were subjected to single

dose pre-treatment via intraperitoneal injection with 400 mg/kg Eucalyptol, where it caused a reduction in NF- κ B expression and, consequently, cytokines and proteinases [47]. Similar results in NF- κ B suppression, compared to Eucalyptol treatment, were also observed in the IPA model caused by cigarette smoke [41] and pneumonia model caused by influenza virus infection (IFV), where BALB / C mice received oral treatment at 120 mg/kg, two days prior to the viral exposure [48]. In addition, Greiner and collaborators [49], suggested a novel mode of NF- κ B blockade through inhibition of nuclear translocation by the nuclear factor kappa B alpha inhibitor (I κ B α) and increased levels in response to treatment with Eucalyptol after stimulation with LPS.

As mentioned above, epithelial cell and macrophage-activated NF- κ B regulate the secretion of many cytokines and chemokines in both asthma and COPD, and these inflammatory mediators play a potential role in the initiation and perpetuation of airway mucus hypersecretion in consequence to inflammatory stimuli [45,50,51].

Under these conditions, cytokines are secreted by the resident tissue cells, and also culminate in the recruitment of leukocytes. Specifically, tumor necrosis factor alpha (TNF- α), interleukin 1b (IL-1 β), interleukin-6 (IL-6), interleukin-8 (IL-8) and interleukin-17 (IL-17) are documented for their important roles in this process and are present in high concentrations in bronchial, lung and sputum biopsy samples of patients with COPD [52-55]. Eucalyptol has been shown to be able to reduce the number of macrophages, as well as the expression of TNF- α , IL-1 β , IL-6 and IL-17, is responsible for the initiation and propagation of inflammation [4,30,36,41,47,48,56].

Another property pertinent to the interaction with these mediators, related to the biological activities of Eucalyptol, relates to an increased expression of interleukin 10 (IL-10) [30,36,48] and cytokine that play an anti-inflammatory role in the innate and adaptive response of the immune system, with significantly lower expression in sputum samples from patients with asthma and COPD [57].

3.4 Cell Adhesion Molecules and Neutrophil Chemotaxis

Some of these cytokines, such as TNF- α factor, IL-1 β , stimulate endothelial cells to express intercellular adhesion molecule (ICAM) -1 and

vascular cell adhesion molecule (VCAM) -1 in bronchial vessels and alveoli, culminating in leukocyte migration to the site of infection [58,59]. In parallel, leukotrienes, a class of eicosanoids present at high levels in asthma and COPD [60], are also capable of inducing the adhesion and activation of leukocytes in the endothelium [61,62]. In contrast to the stimulation of monocytes from asthmatic individuals, Juergens and collaborators [63], observed significant inhibition of cytokines, tramboxane B2 and leukotriene B4 (LTB4) after three days of Eucalyptol therapy with daily doses of 600 mg (3 x 200 mg /day).

Li and collaborators [48], analysed the expression of cell adhesion molecules on the cell surface of mice in response to Influenza virus infection, where positive regulation of ICAM-1 and VCAM-1 was observed, and a significant reduction in the expression of these molecules in the group receiving oral Eucalyptol (120 mg/kg) before and after inoculation of the virus. The results observed in the Eucalyptol treated group, such as suppression of proinflammatory cytokines, transcription factors and adhesion molecules were similar to the positive control group treated with Oseltamivir, the antiviral substance commonly used against influenza virus.

Chemokines, such as IL-8, exert their function by coupling to the G protein of the receptor

expressed in inflammatory cells, regulating their transit towards the pulmonary interstitium [64]. The level of IL-8 is related to the absolute number of neutrophils in induced sputum in individuals with COPD, in addition to being increased in patients with α 1- antiprotease deficiency [44,65,66].

Both *in vitro* [4] and *in vivo* [37] experiments demonstrated the efficacy of Eucalyptol in the inhibition of IL-8, as well as the reduction in the number of leukocytes in bronchoalveolar lavage of mice induced to acute pulmonary inflammation [41].

3.5 Proteases and Remodeling

It is known that neutrophils are implicated in the release of inflammatory cytokines, lipid mediators and enzymes capable of promoting tissue injury [67]. Thus, constant inflammatory stimuli and a growing influx of these leukocytes into the pulmonary parenchyma cause a large release of proteases by these cells, such as matrix metalloproteinases (MMPs). Type 9 matrix metalloprotease (MMP-9) is thought to be the most promising target for drug development due to its predominance in the degrading potential of collagen fibres and elastin, causing pulmonary emphysema and stimulation of mucus hypersecretion, causing chronic bronchitis [19,68-71].

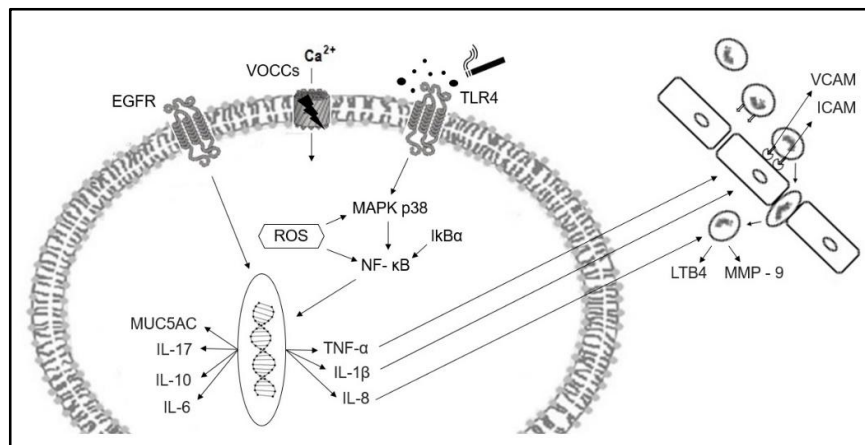


Fig. 1. Schematic diagram, developed from the information collected in the present study, with representation of biological components, which have functionality altered by inhaled irritants (cigarette smoke, air pollutants, indoor dust), capable of interacting with Eucalyptol, such as membrane proteins (EGFR, VOCCs and TLR4), proteins involved in the production of mucus (MUC5AC), elements (TNF- α , IL-1 β , IL-6, IL-8, IL-10, IL-17), transcriptional protein activators (MAP38 p38 and ROS), transcriptional proteins (NF- κ B), leukins (TNF- α , IL-1 β , IL-6, IL-8, IL-10, IL-17), cell adhesion molecules (VCAM, ICAM), LTB4 and MMP-9

In a study conducted by Kim, Lee and Seol [47] it was observed that pre-treatment with Eucalyptol (400 mg/kg), injected intraperitoneally, significantly attenuated the expression of MMP-9 and prevented the histopathological changes caused by said proteolytic enzyme. These results were also similar to the positive control of the study, where dexamethasone was used because it is a drug that has potential anti-inflammatory effects.

Despite having properties that preserve the histoarchitecture of lung tissue, there is a lack in the literature of studies investigating the effects of Eucalyptol in the functional mechanics of the respiratory system. However, Worth et al. [72] conducted a randomised, placebo-controlled study of Eucalyptol (600 mg/kg/day, orally) over 6 months for patients with stable COPD using concomitant pharmacological therapy (β -agonists, anticholinergics and theophylline). In the spirometric protocols, improvement of forced expiratory volume in 1 second (FEV1), vital capacity (CV) and reduction of exacerbations of Eucalyptol-treated group disease in relation to placebo was observed in the spirometric protocols.

The literature review discussed in the present study shows that the biological activities of Eucalyptol when administered orally (100 to 600 mg/kg), intraperitoneal (400 mg/kg), or by inhalation (1 to 10 mg/mL), involve various stages and crucial molecules in the development of the acute and chronic inflammatory process in the respiratory system, as exemplified in Fig.1

4. CONCLUSION

The interaction of Eucalyptol in animal experimental models with pathophysiological mediators (oxidative stress, transcription molecules of cytokines, pro-inflammatory cells and proteases) identified in human respiratory system affections show a relevant alternative treatment option concomitant with the anti-inflammatory drugs in asthma and COPD.

CONSENT

As per international standard or university standard, patient's written consent has been collected and preserved by the authors.

ETHICAL APPROVAL

As per international standard or university standard, written approval of Ethics committee

has been collected and preserved by the authors.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Elwood P, Galante J, Pickering J, Palmer S, Bayer A, Ben-Shlomo Y, Gallacher J. Healthy lifestyles reduce the incidence of chronic diseases and dementia: Evidence from the Caerphilly cohort study. *PLoS one*. 2013;8(12):1-7. doi: 10.1371/journal.pone.0081877
2. Global Strategy for the Diagnosis, Management and Prevention of COPD, Global Initiative for Chronic Obstructive Lung Disease (GOLD); 2017. Available: <http://goldcopd.org>
3. Greiner JFW, Müller J, Zeuner MT, Hauser S, Seidel T, Klenke C, Kaltschmidt B. 1, 8-Cineol inhibits nuclear translocation of NF- κ B p65 and NF- κ B-dependent transcriptional activity. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2013;1833(12):2866-2878. DOI: 10.1016/J.BBAMCR.2013.07.001
4. Juergens UR, Engelen T, Racké K, Stöber M, Gillissen A, Vetter H. Inhibitory activity of 1, 8-cineol (Eucalyptol) on cytokine production in cultured human lymphocytes and monocytes. *Pulmonary pharmacology & therapeutics*. 2004;17(5):281-287. DOI: 10.1016/J.PUPT.2004.06.002
5. Aparicio S, Alcalde R, Dávila MJ, García B, Leal JM. Properties of 1,8-cineole: A thermophysical and theoretical study. *The Journal of Physical Chemistry B*. 2007; 111(12):3167-3177. DOI: 10.1021/jp067405b
6. Liu CH, Chang FY. Development and characterization of Eucalyptol microemulsions for topic delivery of curcumin. *Chemical and Pharmaceutical Bulletin*. 2011;59(2):172-178. DOI: 10.1248/CPB.59.172
7. Barceloux DG. *Medical toxicology of natural substances: Foods, fungi, medicinal herbs, plants, and venomous animals*. John Wiley & Sons; 2012.
8. Ansarin K, Rashidi F, Namdar H, Ghaffari M, Sharifi A. Echocardiographic Evaluation of the Relationship Between inflammatory factors (IL6, TNF α , hs-CRP) and secondary pulmonary hypertension in

- patients with COPD. A cross sectional study. *Pneumologia* (Bucharest, Romania). 2015;64(3):31-35.
9. Brill SE, Law M, El-Emir E, Allinson JP, James P, Maddox V, Nazareth I. Effects of different antibiotic classes on airway bacteria in stable COPD using culture and molecular techniques: A randomised controlled trial. *Thorax*. Thoraxjnl; 2015. DOI: 10.1136/thoraxjnl-2015-207194
 10. Chillappagari S, Preuss J, Licht S, Müller C, Mahavadi P, Sarode G, Henke MO. Altered protease and antiprotease balance during a COPD exacerbation contributes to mucus obstruction. *Respiratory Research*. 2015;16(1):85. doi: 10.1186/s12931-015-0247-x
 11. Esther Jr CR, Coakley RD, Henderson AG, Zhou YH, Wright FA, Boucher RC. Metabolomic evaluation of neutrophilic airway inflammation in cystic fibrosis. *CHEST Journal*. 2015;148(2):507-515. DOI: 10.1378/chest.14-1800
 12. Gane JM, Stockley RA, Sapey E. The rs361525 polymorphism does not increase production of tumor necrosis factor alpha by monocytes from alpha-1 antitrypsin deficient subjects with chronic obstructive pulmonary disease-a pilot study. *Journal of Negative Results in Biomedicine*. 2015; 14(1):20. DOI: 10.1186/s12952-015-0039-3
 13. Wells JM, Jackson PL, Viera L, Bhatt SP, Gautney J, Handley G, Dransfield MT. A randomized, placebo-controlled trial of roflumilast. Effect on proline-glycine-proline and neutrophilic inflammation in chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*. 2015;192(8):934-942. DOI: 10.1164/rccm.201503-0543oc
 14. YUN CM, SANG XY. Role of proteinase-activated receptor-1 gene polymorphisms in susceptibility to chronic obstructive pulmonary disease. *Genetics and Molecular Research*. 2015;14(4):13215-13220. DOI: 10.4238/2015.October.26.18
 15. Cortez-Lugo M, Ramírez-Aguilar M, Pérez-Padilla, R, Sansores-Martínez R, Ramírez-Venegas A, Barraza-Villarreal A. Effect of personal exposure to PM2.5 on respiratory health in a Mexican panel of patients with COPD. *International Journal of Environmental Research and Public Health*. 2015;12(9):10635-10647. DOI: 10.3390/ijerph120910635
 16. Minakata Y, Morishita Y, Ichikawa T, Akamatsu K, Hirano T, Nakanishi M, Ichinose M. Effects of pharmacologic treatment based on airflow limitation and breathlessness on daily physical activity in patients with chronic obstructive pulmonary disease. *International Journal of Chronic Obstructive Pulmonary Disease*. 2015;10: 1275. DOI: 10.2147/COPD.S84134
 17. Topalovic M, Derom E, Osadnik CR, Troosters T, Decramer M, Janssens W. Airways resistance and specific conductance for the diagnosis of obstructive airways diseases. *Respiratory Research*. 2015;16(1):88. DOI: 10.1186/S12931-015-0252-0
 18. Wei J, Zhao H, Fan G, Li J. Bilirubin treatment suppresses pulmonary inflammation in a rat model of smoke-induced emphysema. *Biochemical and Biophysical Research Communications*. 2015;465(2):180-187. DOI: 10.1016/j.bbrc.2015.07.133
 19. Barnes PJ. New anti-inflammatory targets for chronic obstructive pulmonary disease. *Nature Reviews Drug Discovery*. 2013;12(7):543-559. DOI: 10.1038/nrd4025
 20. Global initiative for chronic obstructive lung disease. Global strategy for the diagnosis, management, and prevention of copd; 2017. Available:<http://goldcopd.org/global-strategy-diagnosis-management-prevention-copd-2016/>
 21. Burgel PR, Nadel JA. Epidermal growth factor receptor-mediated innate immune responses and their roles in airway diseases. *European Respiratory Journal*. 2008;32(4):1068-1081. DOI: 10.1183/09031936.00172007
 22. Calafat J, Janssen H, Zuurbier AE, Knol EF, Egesten A. Human monocytes and neutrophils store transforming growth factor- α in a subpopulation of cytoplasmic granules. *Blood*. 1997;90(3):1255-1266.
 23. Rumelhard M, Ramgolam K, Hamel R, Marano F, Baeza-Squiban A. Expression and role of EGFR ligands induced in airway cells by PM2.5 and its components. *European Respiratory Journal*. 2007;30(6):1064-1073. DOI: 10.1183/09031936.00085907
 24. Takeyama K, Fahy JV, Nadel JA. Relationship of epidermal growth factor

- receptors to goblet cell production in human bronchi. *American Journal of Respiratory and Critical Care Medicine*. 2001;163(2):511-516.
DOI: 10.1164/AJRCCM.163.2.2001038
25. O'donnell RA, Richter A, Ward J, Angco G, Mehta A, Rousseau K, Wilson SJ. Expression of ErbB receptors and mucins in the airways of long term current smokers. *Thorax*. 2004;59(12):1032-1040.
DOI: 10.1136/THX.2004.028043
26. Liu L, Tsa, JC, Aird, William C. Egr-1 gene is induced by the systemic administration of the vascular endothelial growth factor and the epidermal growth factor. *Blood*. 2000;96(5):1772-1781.
27. Cho SJ, Kang MJ, Homer RJ, Kang HR, Zhang X, Lee PJ, Lee CG. Role of early growth response-1 (Egr-1) in interleukin-13-induced inflammation and remodeling. *Journal of Biological Chemistry*. 2006;281(12):8161-8168.
28. Zhou X, Dai Q, Huang X. Neutrophils in acute lung injury. *Frontiers in bioscience (Landmark edition)*. 2011;17:2278-2283.
29. Nascimento NRF, Refosco RMD, Vasconcelos ECF, Kerntopf MR, Santos CF, Batista, FJA, Fonteles MC. 1, 8-Cineole induces relaxation in rat and guinea-pig airway smooth muscle. *Journal of Pharmacy and Pharmacology*. 2009; 61(3):361-366.
DOI: 10.1211/JPP.61.03.0011
30. Bastos VP, Gomes AS, Lima FJ, Brito TS, Soares PM, Pinho JP, Magalhães PJ. Inhaled 1, 8-cineole reduces inflammatory parameters in airways of Ovalbumin-challenged Guinea Pigs. *Basic & Clinical Pharmacology & Toxicology*. 2011;108(1):34-39.
DOI: 10.1111/J.1742-7843.2010.00622.X
31. Soares MCMS, Damiani CEN, Moreira CM, Stefanon I, Vassallo DV. Eucalyptol, an essential oil, reduces contractile activity in rat cardiac muscle. *Brazilian Journal of Medical and Biological Research*. 2005; 38(3):453-461.
DOI: 10.1590/S0100-79X2005000300017
32. Jarvis D, Zock JP, Heinrich J, Svanes C, Verlato G, Olivieri M, Dahlman-Hoglund A. Cat and dust mite allergen levels, specific IgG and IgG 4 and respiratory symptoms in adults. *Journal of allergy and clinical immunology*. 2007;119(3):697-704.
DOI: 10.1016/J.JACI.2006.10.042
33. Freeman CM, Martinez FJ, Han MK, Washko GR, McCubbrey AL, Chensue SW, Curtis JL. Lung CD8+ T cells in COPD have increased expression of bacterial TLRs. *Respiratory Research*. 2013;14(1): 13.
DOI: 10.1186/1465-9921-14-13
34. Nadigel J, Préfontaine D, Baglolle CJ, Maltais F, Bourbeau J, Eidelman DH, Hamid Q. Cigarette smoke increases TLR4 and TLR9 expression and induces cytokine production from CD8+ T cells in chronic obstructive pulmonary disease. *Respiratory Research*. 2011;12(1):149.
DOI: 10.1186/1465-9921-12-149
35. Vassallo R, Walters PR, Lamont J, Kottom TJ, Eunhee SY, Limper AH. Cigarette smoke promotes dendritic cell accumulation in COPD; a Lung Tissue Research Consortium study. *Respiratory Research*. 2010;11(1):45.
DOI: 10.1186/1465-9921-11-45
36. Zhao C, Sun J, Fang C, Tang F. 1, 8-cineol attenuates LPS-induced acute pulmonary inflammation in mice. *Inflammation*. 2014; 37(2):566-572.
DOI: 10.1007/s10753-013-9770-4
37. Lee HS, Park DE, Song WJ, Park HW, Kang HR, Cho SH, Sohn SW. Effect of 1,8-cineole in dermatophagoides pteronyssinus-stimulated bronchial epithelial cells and mouse model of asthma. *Biological and Pharmaceutical Bulletin*. 2016;39(6):946-952.
DOI: 10.1248/BPB.B15-00876
38. Kirkham PA, Caramori G, Casolari P, Papi AA, Edwards M, Shamji B, Heinemann L. Oxidative stress-induced antibodies to carbonyl-modified protein correlate with severity of chronic obstructive pulmonary disease. *American journal of Respiratory and Critical Care Medicine*. 2011;184(7): 796-802.
DOI: 10.1164/Rccm.201010-1605oc
39. Pourazar J, Blomberg A, Kelly FJ, Davies DE, Wilson SJ, Holgate ST, Sandström, T. Diesel exhaust increases EGFR and phosphorylated cterminal Tyr 1173 in the bronchial epithelium. *Particle and Fibre Toxicology*. 2008;5(1):1-9.
DOI: 10.1186/1743-8977-5-8
40. Rahman I, Biswas SK, Jimenez LA, Torres M, Forman HJ. Glutathione, stress responses, and redox signaling in lung inflammation. *Antioxidants & Redox Signaling*. 2005;7(1-2):42-59.
DOI: 10.1089/ARS.2005.7.42
41. Kennedy-Feitosa E, Okuro, RT, Ribeiro VP, Lanzetti M, Barroso MV, Zin, WA,

- Valenca SS. Eucalyptol attenuates cigarette smoke-induced acute lung inflammation and oxidative stress in the mouse. *Pulmonary Pharmacology & Therapeutics*. 2016;41:11-18.
DOI: 10.1016/J.PUPT.2016.09.004
42. Carpentier I, Declercq W, Malinin NL, Wallach D, Fiers W, Beyaert R. TRAF2 plays a dual role in NF- κ B-dependent gene activation by mediating the TNF-induced activation of p38 MAPK and I κ B kinase pathways. *Febs Letters*. 1998;425(2):195-198.
DOI: 10.1016/S0014-5793(98)00226-9
43. Saatian B, Yutong ZY, He D, Georas SN, Watkins T, Spannhake E, Natarajan V. Transcriptional regulation of lysophosphatidic acid-induced interleukin-8 expression and secretion by p38 MAPK and JNK in human bronchial epithelial cells. *Biochemical Journal*. 2006;393(3): 657-668.
DOI: 10.1042/bj20050791
44. Pourazar J, Mudway IS, Samet JM, Helleday R, Blomberg A, Wilson SJ, Sandstrom T. Diesel exhaust activates redox-sensitive transcription factors and kinases in human airways. *American Journal of Physiology-Lung Cellular and Molecular Physiology*. 2005;289(5):L724-L730.
DOI: 10.1152/AJPLUNG.00055.2005
45. Khansari N, Shakiba Y, Mahmoudi M. Chronic inflammation and oxidative stress as a major cause of age-related diseases and cancer. *Recent Patents on Inflammation & Allergy Drug Discovery*. 2009;3(1):73-80.
DOI: 10.2174/187221309787158371
46. Karin M, Yamamoto Y, Wang QM. The IKK NF- κ B system: A treasure trove for drug development. *Nature Reviews Drug Discovery*. 2004;3(1):17-26.
DOI: 10.1038/NRD1279
47. Kim KY, Lee HS, Seol GH. Eucalyptol suppresses matrix metalloproteinase-9 expression through an extracellular signal-regulated kinase-dependent nuclear factor-kappa B pathway to exert anti-inflammatory effects in an acute lung inflammation model. *Journal of Pharmacy and Pharmacology*. 2015;67(8):1066-1074.
DOI: 10.1111/JPHP.12407
48. Li, Y, Lai Y, Wang Y, Liu N, Zhang F, Xu P. 1, 8-Cineol protect against influenza-virus-induced pneumonia in mice. *Inflammation*. 2016;39(4):1582-1593.
DOI: 10.1007/s10753-016-0394-3
49. Greiner JFW, Müller J, Zeuner MT, Hauser S, Seidel T, Klenke C, Kaltschmidt B. 1, 8-Cineol inhibits nuclear translocation of NF- κ B p65 and NF- κ B-dependent transcriptional activity. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2013;1833(12):2866-2878.
DOI: 10.1016/J.BBAMCR.2013.07.001
50. Lundgren JD, Shelhamer JH. Pathogenesis of airway mucus hypersecretion. *Journal of allergy and clinical immunology*. 1990;85(2):399-417.
DOI: 10.1016/0091-6749(90)90147-V
51. Caramori G, Casolari P, Adcock I. Role of transcription factors in the pathogenesis of asthma and COPD. *Cell Communication & Adhesion*. 2013;20(1-2):21-40.
DOI: 10.3109/141961.2013.775257
52. Ricci M, Matucci A, Rossi O. Recent advances in the pathogenetic mechanisms and genetic aspects of atopic diseases. *Allergy Clin Immunol News*. 1994;6:103-8.
53. Di Stefano A, Caramori G, Gnemmi I, Contoli M, Vicari C, Capelli A, Casolari P. T helper type 17-related cytokine expression is increased in the bronchial mucosa of stable chronic obstructive pulmonary disease patients. *Clinical & Experimental Immunology*. 2009;157(2): 316-324.
DOI: 10.1111/j.1365-2249.2009.03965.x
54. Pridgeon C, Bugeon L, Donnelly L, Straschil U, Tudhope SJ, Fenwick P, Dallman MJ. Regulation of IL-17 in chronic inflammation in the human lung. *Clinical Science*. 2011;120(12):515-524.
DOI: 10.1042/cs20100417
55. Marumo S, Hoshino Y, Kiyokawa H, Tanabe N, Sato A, Ogawa E, Mishima M. p38 mitogen-activated protein kinase determines the susceptibility to cigarette smoke-induced emphysema in mice. *BMC pulmonary medicine*. 2014;14(1):79.
DOI: 10.1186/1471-2466-14-79
56. Sadlon AE, Lamson DW. Immune-modifying and antimicrobial effects of Eucalyptus oil and simple inhalation devices. *Alternative Medicine Review*. 2010;15(1):33-43.
57. Takanashi S, Hasegawa Y, Kanehira Y, Yamamoto K, Fujimoto K, Satoh K, Okamura K. Interleukin-10 level in sputum is reduced in bronchial asthma, COPD and in smokers. *European Respiratory Journal*. 1999;14(2):309-314.

58. Tosi MF, Stark JM, Smith C, Hamedani A, Gruenert D, Infeld MD. Induction of ICAM-1 expression on human airway epithelial cells by inflammatory cytokines: Effect of neutrophil-epithelial cells adhesion. *Am J Respir Cell Mol Biol.* 1992;7:214-21. DOI: 10.1165/ajrcmb/7.2.214
59. Kumar V, Cotran RS, Robbins SL. *Patologia humana.* Elsevier Health Sciences; 2008.
60. Boyce JA. Eicosanoids in asthma, allergic inflammation, and host defense. *Current molecular medicine.* 2008;8(5): 335-349. DOI: 10.2174/156652408785160989
61. Goetzi EJ, PICKETT WC. The human PMN leukocyte chemotactic activity of complex hydroxy-eicosatetraenoic acids (HETEs). *The Journal of Immunology.* 1980;125(4):1789-1791.
62. Mayes PA, Botham KM. Metabolism of unsaturated fatty acids and eicosanoids. *Harper's Illustrated Biochemistry, 26th Ed.* (Lange Medical Books, New York). 2003; 190-196.
63. Juergens UR, Stöber M, Vetter H. Inhibition of cytokine production and arachidonic acid metabolism by Eucalyptol (1,8-cineole) in human blood monocytes in vitro. *European Journal of Medical Research.* 1998;3(11):508-510.
64. Yamagata T, Ichinose M. Agents against cytokine synthesis or receptors. *European Journal of Pharmacology.* 2006;533(1): 289-301. DOI: 10.1016/j.ejphar.2005.12.046
65. Woolhouse IS, Bayley DL, Stockley RA. Sputum chemotactic activity in chronic obstructive pulmonary disease: Effect of α 1-antitrypsin deficiency and the role of leukotriene B4 and interleukin 8. *Thorax.* 2002;57(8):709-714. DOI: 10.1136/THORAX.57.8.709
66. Rufino R, Costa CHD, Souza HSPD, Madi K, Silva JRL. Induced sputum and peripheral blood cell profile in chronic obstructive pulmonary disease. *Journal Brasileiro de Pneumologia.* 2007;33(5): 510-518. DOI: 10.1590/S1806-37132007000500005
67. Holz O, Seiler T, Karmeier A, Fraedrich J, Leiner H, Magnussen H, Welker L. Assessing airway inflammation in clinical practice—experience with spontaneous sputum analysis. *BMC Pulmonary Medicine.* 2008;8(1):5. DOI: 10.1186/1471-2466-8-5
68. Parks WC, Wilson CL, López-Boado YS. Matrix metalloproteinases as modulators of inflammation and innate immunity. *Nature Reviews Immunology.* 2004;4(8):617-629. DOI: 10.1038/NRI1418
69. Macnee W. Pathogenesis of chronic obstructive pulmonary disease. *Proceedings of the American Thoracic Society.* 2005;2(4):258-266. DOI: 10.1513/pats.200504-045SR
70. Valença SS, Porto LC. Immunohistochemical study of lung remodeling in mice exposed to cigarette smoke. *Journal Brasileiro de Pneumologia.* 2008;34(10): 787-795. DOI: 10.190/S1806-37132008001000006
71. Grzela K, Litwiniuk M, Zagorska W, Grzela T. Airway remodeling in chronic obstructive pulmonary disease and asthma: The role of matrix metalloproteinase-9. *Archivum Immunologiae et Therapiae Experimentalis.* 2016;64(1):47-55. DOI: 10.1007/s00005-015-0345-y
72. Worth H, Schacher C, Dethlefsen U. Concomitant therapy with Cineole (Eucalyptole) reduces exacerbations in COPD: A placebo-controlled double-blind trial. *Respiratory Research.* 2009;10(1):69. DOI: 10.1186/1465-9921-10-69

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